

# Immunohistochemical Identification And Quantification Of Brown Adipocytes In Human Adipose Tissue.

\* Insaf Jasim Mahmoud, MBChB, PhD, \*Abdul Hadi Liebi MBChB, CABS , \*Muthanna Alasal, MBChB, FICMS,

\*Faris Abdul Kareem Khazaal, MBChB, CABM, FRCP

## ABSTRACT

**Background:** Obesity is a worldwide challenge and is closely connected to many metabolic diseases. Two types of adipose tissue, white adipose tissue (WAT) and brown adipose tissue (BAT) have been identified. White fat cells store chemical energy, brown adipocytes defend against hypothermia, obesity and diabetes.

**Objective:** To localize and quantify brown adipocytes in human subcutaneous (S) and visceral (V) adipose tissue by histology and immunohistochemistry.

**Type of the study:** A cross-sectional study.

**Methods:** Adipose tissue was obtained from histopathology specimens taken from ten patients, of different age, sex and body mass index (BMI), undergoing surgery for different pathologies. Immunohistochemistry for detection of UCP (Uncoupling Protein) in S and V adipose tissue depots was performed, and percentage of the positive pixels for UCP color intensity was measured and statistical analysis performed. data was expressed as mean  $\pm$  standard deviation (SD) and analyzed using student t-test to compare values

**Results:** Brown adipocytes with typical multivesicular appearance were detected as clusters of cells among white fat in both S and V adipose tissue. The percentage of the positive pixels for UCP color intensity of brown adipocytes was significantly ( $P < 0.001$  and  $P < 0.05$ ) higher in S than in V adipose tissue, significantly higher levels ( $P < 0.05$ ) when BMI was 30 or more, and non-significant higher levels ( $P > 0.05$ ) in females than males.

**Conclusion:** Brown adipocytes are more abundant in S than V adipose tissue, and have a positive correlation to BMI.

**Keywords:** human brown adipocytes, immunohistochemistry, UCP.

*Al-Kindy College Medical Journal 2016: Vol. 12 No.2  
Page: 84-88*

\* Obesity research and therapy unit / Alkindy College of Medicine

*Received 8<sup>th</sup> Feb 2016, accepted in final 3<sup>rd</sup> Oct 2016*

*Corresponding to :* Insaf Jasim Mahmoud email: insafh@yahoo.com

Obesity is a worldwide challenge and not unique to anyone country. Furthermore, obesity is closely connected to many metabolic diseases. Essentially, obesity and overweight are caused by the energy imbalance between the calories consumed and calories expended.

Adipose tissue, which is composed mostly of adipocytes, is a major endocrine organ and plays a key role in energy homeostasis. Two types of adipose tissue, white adipose tissue (WAT) and brown adipose tissue (BAT), have been identified (1)

While white fat cells are specialized to store chemical energy, brown adipocytes defend mammals against hypothermia, obesity and diabetes. Brown fat utilizes a high mitochondrial content and high mitochondrial uncoupling protein one (UCP1) to uncouple respiration and dissipate chemical energy as heat. Rodents and other small mammals have copious brown fat deposits, but larger mammals often lose prominent brown fat depots after infancy. (2, 3, 4, 5, 6)

The physiological significance of adult human brown fat has not yet been fully explored. It has been known for many years that some white adipose tissues contain cells that can express high levels of UCP1 and take on a

multilocular appearance upon prolonged stimulation by cold or pathways that elevate intracellular cyclic AMP. (7, 8)

In humans and other large mammalian species, BAT was traditionally thought to be restricted to the neonatal and early childhood periods (9, 10). However, positron emission tomography (PET) scanning technology was recently adopted for detecting metabolically active sites for oncology diagnosis; this application is based on the uptake of radio labeled non-metabolizable glucose derivatives. The results obtained from a scanning experiment using PET to analyze BAT clearly demonstrated that active BAT is present in adult humans at discrete anatomical sites, especially in the upper trunk, such as cervical, supraclavicular, paravertebral, pericardial, and to some extent, mediastinal and mesenteric areas. (2, 5, 11, 12)

Recently, a new type of brown-like adipocyte was discovered that shows distinct gene expression patterns from those of white or brown adipocytes. These novel brown-like cells that reside within WAT, especially inguinal WAT, were termed beige/brite adipocytes. (13) Using Heaton's definition of BAT as fat containing multilocular adipocytes stained by hematoxylin-eosin on

light microscopy (14)BAT was designated as being visceral (V) or subcutaneous (S), subdividing each category into separate depots according to their contiguous organ or tissue. (11, 15) In healthy lean male and female adults, age  $40 \pm 9$  years, mean weight of cold-activated human BAT calculated by PET-CT was 34 g (range 9-90) (4). Based on the calculation of 63 g cervical BAT reported in a different study by Virtanen et al. (12) it was estimated that activated BAT thermogenesis contributed 4.5% to whole body energy expenditure, which was considered to be significant (16) and commensurate with the goal of exploiting BAT to burn excess calories stored in WAT for weight loss in obesity and type 2 diabetes (15, 17). The 5% figure cited above could be an underestimate by a factor of two to three in some subjects because of their greater BAT mass.

More extensive analysis was necessary in human BAT studies, thus we performed this work to localize and quantify brown adipocytes in human adipose tissue by histology and immunohistochemistry.

**Methods:** Adipose tissue was obtained from histopathology specimens taken from patients undergoing surgery for trauma to chest or abdomen in Al Kindy Teaching Hospital in Baghdad. These patients were ten in number and of different age, sex and body mass index (BMI). Study protocol was approved by scientific and ethical committee in Al Kindy College of Medicine.

**Immunohistochemistry:** for detection of UCP in adipose tissue, 4 $\mu$ m paraffin sections were mounted on Fisher brand positively charged slides (18). Actual assessment of immunohistochemistry was performed by image analysis of tissue sections using Aperio image scope v11.1.2.760 software program and positive pixel count algorithms were used to quantify the amount of a specific color in a slide image. This algorithm has a set of default input parameters when first selected. These inputs were pre-configured for brown color quantification in the three intensity ranges (weak, positive, and strong).

Immunohistochemistry staining in adipose tissue sections were quantified as the percentage of the positive pixels for UCP color intensity. The data was expressed as mean  $\pm$  standard deviation (SD) and analyzed using student t-test to compare values.

Differences between groups were considered highly significant at ( $P < 0.001$ ), significant at ( $P < 0.05$ ) or non-significant at ( $P > 0.05$ ).

**Results :** Brown adipocytes were detected as clusters of cells among white fat cells (Figure 2). They were found in both subcutaneous and visceral adipose tissue that was collected from different body regions (Figure a, Figure b). At a higher magnification, the brown adipocytes showed the typical appearance of multi vesicle - filled cytoplasm (Figure 3). The percentage of the positive pixels for UCP color intensity of brown adipocytes detected by immunohistochemistry staining in adipose tissue sections showed that there was

significant higher levels in subcutaneous than in visceral adipose tissue regions (Table 1). The percentage of positive pixels for brown adipocytes in adipose tissue with respect to BMI showed significantly higher levels ( $P < 0.05$ ) when BMI was 30 or more (Table 2). Concerning gender, there was a non-significant higher level in the percentage of positive pixels for brown adipocytes in females than males ( $P > 0.05$ ) (Table-3).

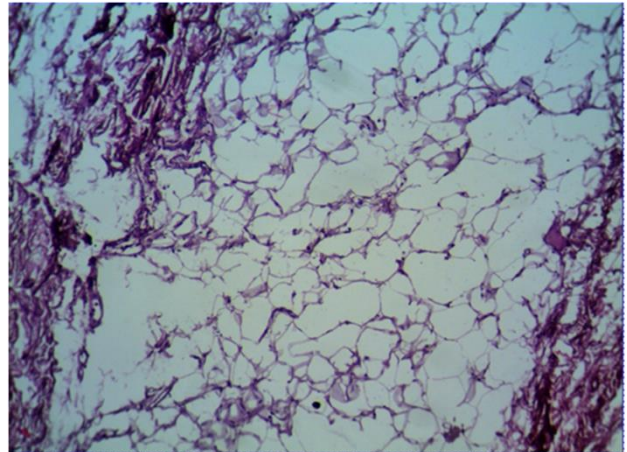


Figure 1- Subcutaneous adipose tissue. Hematoxylin and Eosin, 100x.

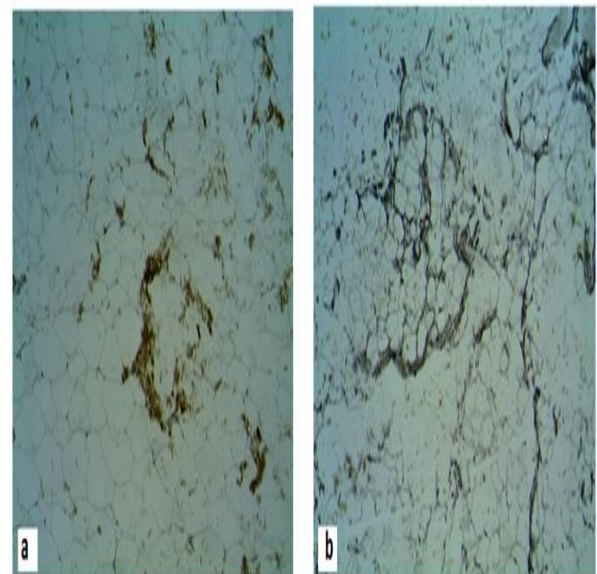


Figure 2- Immunohistochemistry for UCP identification in brown adipocytes. a. subcutaneous adipose tissue b. visceral adipose tissue. Staining by peroxidase/DAB (brown).100x.

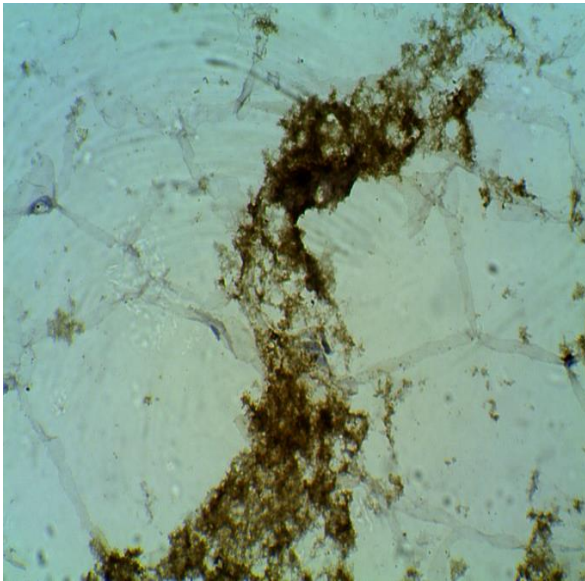


Figure 3- Immunohistochemistry for UCP identification in brown adipocytes (subcutaneous) showing multilocular appearance. Staining by peroxidase/DAB (brown).400x.

Male	Female	
Brown adipocytes Mean ± SD	Brown adipocytes Mean ± SD	P value
22.7 ± 14.8	28.8 ± 17.6	0.124

**Table-3:** Percentage of positive pixels for brown adipocytes in adipose tissue with respect to gender. (P > 0.05): non- significant

**Discussion :**The ability of brown fat to suppress obesity through increased energy expenditure has caused an explosion of interest in the development and function of brown adipocytes (17). With the use of different techniques, many studies have been performed to identify and localize brown adipose tissue (2, 5, 11 and 12), and brown adipocytes within white adipose tissue (13), however none has aimed to quantify the number and distribution of brown adipocytes among white fat cells using immunohistochemical technique in human adipose tissue depots. At present all known adipose-specific genes are expressed in both white and brown adipocytes (adipocyte lipid binding protein, adiponin, etc.) or in brown adipocytes only (UCP), with none being expressed exclusively in white adipocytes (20, 21, 22 and 23). Thus the demonstration of brown color intensity using immunohistochemistry for UCP would indicate brown adipocytes.

Our finding that there is significant difference in number of brown adipocytes, reflected by the percent of positive pixels of brown color, in subcutaneous and visceral adipose tissues, being more in the subcutaneous region may be due to the presence of a greater source of precursor cells for brown adipocytes in the subcutaneous region which are myf-5, muscle-like cellular lineage (24); or from transdifferentiation from mature white adipocytes into beige (brite) adipocytes (25) or may be due to the presence of distinct genetic loci that control the amounts of UCP1-positive cells in the white and classical brown fat depots (24, 26, 27, 28 and 29) and that these loci are present more in the subcutaneous adipose tissue depots (30). We found a positive relation between the quantity of brown adipocytes and BMI. This result is in contrast to what has been found by others who found a negative correlation between body fat and brown adipose tissue (31, 32). However, all these studies were correlating brown adipose tissue and different functional parameters as glucose metabolism (31), exposure to cold, age, sex and lipid profile (32). Our results showed non- significant higher levels of brown adipocytes in females than males. This finding was similar to what has been found in one study (32), where there was a significant sexual difference (5.5% in the females vs 1.3% in the males; P<0.000).

	S Brown adipocytes Mean ± SD	V Brown adipocytes Mean ± SD	p values
s1 vs v1	31.97 ± 3.05	21.25 ± 2.84	0.004
s2 vs v2	25.77 ± 2.80	17.84 ± 2.49	0.008
s3 vs v3	25.64 ± 9.17	24.5 ± 14.1	0.896
s4 vs v4	20.86 ± 8.12	5.330 ± 0.618	0.032
s5 vs v5	36.62 ± 7.95	43.70 ± 5.17	0.196
s6 vs v6	24.73 ± 1.06	5.8315 ± 0.0721	0.000
s7 vs v7	44.6 ± 15.7	11.91 ± 6.20	0.030
s8 vs v8	58.52 ± 9.03	11.91 ± 6.20	0.000
s9 vs v9	33.70 ± 2.26	37.1 ± 36.7	0.864

**Table-1:** Percentage of positive pixels for brown adipocytes in subcutaneous (S) versus visceral (V) adipose tissue. (No. 1,2,3..etc. referred to the no. of patient)

(P < 0.001): highly significant  
(P < 0.05): significant  
(P > 0.05): non- significant

BMI <30	BMI ≥30	
Brown adipocytes Mean ± SD	Brown adipocytes Mean ± SD	P value
20.50 ± 9.05	29.6 ± 18.4	0.005

**Table-2:** Percentage of positive pixels for brown adipocytes in adipose tissue with respect to BMI. (P < 0.05): significant



## Conclusion

Brown and browning Adipose tissue are more abundant in subcutaneous than visceral adipose tissue and there is positive correlation between brown adipocytes and body mass index.

## References

1. Bae KH, Kim WK, Lee SC. Involvement of protein tyrosine phosphatases in adipogenesis: new anti-obesity targets? *BMB Rep* 2012; 45 : 700-706
2. Cypess AM , Lehman S, Williams G, Tal I, Rodman D, Gold - fine AB, Kuo FC, Palmer EL, Tseng YH, Doria A, Kolodny GM, Kahn CR. Identification and importance of brown adipose tissue in adult humans. *N Engl J Med* 2009; 360 : 1509-1517
3. Mirbolooki MR, Constantinescu CC, Pan ML, Mukherjee J. Quantitative assessment of brown adipose tissue metabolic activity and volume using 18F-FDG PET/CT and  $\beta$ 3-adrenergic receptor activation. *EJNMMI Res.* 2011; 1:30.
4. Orava J, Nuutila P, Lidell ME, Oikonen V, Nojonen T, Viljanen T, Scheinin M, Taittonen M, Niemi T, Enerback S, et al. Different metabolic responses of human brown adipose tissue to activation by cold and insulin. *Cell Metab.* 2011; 14:272-279.
5. van Marken Lichtenbelt WD, Vanhommerig JW, Smulders NM, Drossaerts JM, Kemerink GJ, Bouvy ND, Schrauwen P, Teule GJ. Cold-activated brown adipose tissue in healthy men. *N Engl J Med.* 2009; 360:1500-1508.
6. Virtanen KA, Lidell ME, Orava J, Heglind M, Westergren R, Niemi T, Taittonen M, Laine J, Savisto NJ, Enerback S, et al. Functional brown adipose tissue in healthy adults. *N Engl J Med.* 2009; 360:1518-1525
7. Cousin B, Cinti S, Morrioni M, Raimbault S, Ricquier D, Penicaud L, Casteilla L. Occurrence of brown adipocytes in rat white adipose tissue: molecular and morphological characterization. *J Cell Sci.* 1992; 103:931-942.
8. Young P, Arch JR, Ashwell M. Brown adipose tissue in the parametrial fat pad of the mouse. *FEBS Lett.* 1984; 167:10-14.
9. Lean ME. Brown adipose tissue in humans. *Proc Nutr Soc* 1989; 48 : 243-256
10. Enerbäck S. Human brown adipose tissue. *Cell Metab* 2010; 11: 248-252
11. Nedergaard J, Bengtsson T, Cannon B. Unexpected evidence for active brown adipose tissue in adult humans. *Am J Physiol Endocrinol Metab* 2007; 293
12. Virtanen KA , Lidell ME, Orava J, Heglind M, Westergren R, Niemi T, Taittonen M, Laine J, Savisto NJ, Enerbäck S, Nuutila P. Functional brown adipose tissue in healthy adults. *N Engl J Med* 2009; 360: 1518-1525
13. Jun Wu<sup>1</sup>, Pontus Boström<sup>1</sup>, Lauren M. Sparks<sup>2</sup>, Li Ye<sup>1</sup>, Jang Hyun Choi<sup>1</sup>, An-Hoa Giang, Melin Khandekar, Pirjo Nuutila, Gert Schaart, Kexin Huang, Hua Tu, Wouter D. van Marken Lichtenbelt, Joris Hoeks, Sven Enerbäck, Patrick Schrauwen, and Bruce M. Spiegelman. Beige Adipocytes are a Distinct Type of Thermogenic Fat Cell in Mouse and Human Cell. 2012 July 20; 150 (2): 366-376
14. Heaton JM. The distribution of brown adipose tissue in the human. *J Anat* 1972; 112: 35-39.
15. Harold Sacks and Michael Symonds. Anatomical Locations of Human Brown Adipose Tissue Functional Relevance and Implications in Obesity and Type 2 Diabetes. *Diabetes* 2013; 62: 1783-1790.
16. van Marken Lichtenbelt WD, Schrauwen P. Implications of nonshivering thermogenesis for energy balance regulation in humans. *Am J Physiol Regul Integr Comp Physiol* 2011; 301: R285-R296.
17. Cypess AM, Kahn CR. Brown fat as a therapy for obesity and diabetes. *Curr Opin Endocrinol Diabetes Obes* 2010; 17: 143-149.
18. Bancroft JD and Marilyn Gamble. Tissue processing. In: Theory and practice of histology techniques. Bancroft, J.D. (Ed), 6th edition. Churchill Livingstone. Elsevier. 2008; 21: pp.433-472.
19. Saverio Cinti, Robert C. Frederich, M. Cristina Zingaretti, Rita de Matteis, Jjeffrey S. Flier, and Bradford B. Lowell. Immunohistochemical localization of leptin and uncoupling protein in white and brown adipose tissue. *Endocrinology* 1997; 138: 797-804.
20. Ailhaud G, Grimaldi P, Negrel R. Cellular and molecular aspects of adipose tissue development. *Annu Rev Nutr* 1992; 12: 207-233
21. Spiegelman BM, Choy L, Hotamisligil GS, Graves RS, Tontonoz P. Regulation of adipocyte gene expression in differentiation and syndromes of obesity/diabetes. *J Biol Chem* 1993; 268: 6823-6826.
22. Ricquier D, Cassard-Doulcier AM. The biochemistry of white and brown adipocytes analyzed from a selection of proteins. *Eur J Biochem* 1993; 218: 785- 796.
23. Himms-Hagen J, Melnyk A, Zingaretti MC, Ceresi E, Barbatelli G, Cinti S. Multilocular fat cells in WAT of CL-316243-treated rats derive directly from white adipocytes. *Am J Physiol Cell Physiol.* 2000; 279: C670-681.
24. Seale P, Bjork B, Yang W, Kajimura S, Chin S, Kuang S, Scime A, Devarakonda S, Conroe HM, Erdjument-Bromage H, et al. PRDM16

- controls a brown fat/skeletal muscle switch. *Nature*. 2008; 454:961-967.
25. Shingo Kajimura and Masayuki Saito. A new era in brown adipose tissue biology: molecular control of brown fat development and energy homeostasis. *Annu. Rev. Physiol.* 2014; 76:13.1-13.25.
  26. Coulter AA, Bearden CM, Liu X, Koza RA, Kozak LP. Dietary fat interacts with QTLs controlling induction of Pgc-1 alpha and Ucp1 during conversion of white to brown fat. *Physiol Genomics*. 2003; 14:139-147
  27. Guerra C, Koza RA, Yamashita H, Walsh K, Kozak LP. Emergence of brown adipocytes in white fat in mice is under genetic control. Effects on body weight and adiposity. *J Clin Invest*. 1998; 102:412-420.
  28. Koza RA, Hohmann SM, Guerra C, Rossmeisl M, Kozak LP. Synergistic gene interactions control the induction of the mitochondrial uncoupling protein (Ucp1) gene in white fat tissue. *J Biol Chem* 2000; 275:34486-34492.
  29. Xue B, Coulter A, Rim JS, Koza RA, Kozak LP. Transcriptional synergy and the regulation of Ucp1 during brown adipocyte induction in white fat depots. *Mol Cell Biol*. 2005; 25:8311-8322.
  30. Xue B, Rim JS, Hogan JC, Coulter AA, Koza RA, Kozak LP. Genetic variability affects the development of brown adipocytes in white fat but not in interscapular brown fat. *J Lipid Res*. 2007; 48:41-51.
  31. M Matsushita, T Yoneshiro, S Aita, T Kameya, H Sugie and M Saito. Impact of brown adipose tissue on body fatness and glucose metabolism in healthy humans. *Int J Obes (Lond)*. 2014 ; 38(6):812-7
  32. W. Wang, Q. Wang, M. Zhang, M. Xu, W. Gu, L. Qi, B. Li & G. Ning. Brown adipose tissue activation is inversely related with central obesity and metabolic parameters in adult human. *Endocrine Abstracts* 2012; 29.