

Al-Kindy College Medical Journal (KCMJ)

Review Article

Melatonin in male and female fertility

Zainab M. Alawad^{*}, Hanan L. Al-Omary

Department of Physiology, College of Medicine, University of Baghdad, Baghdad. Iraq

ABSTRACT

* Corresponding author: <u>zainabm.alawad@comed.uobaghdad.edu.iq</u>

Article history: Received 28 July 2021 Accepted 27 September 2021 Available online 30 December 2021

https://doi.org/10.47723/kcmj.v17i3.398

Keywords: Melatonin, Antioxidant, Male fertility, Female fertility, In Vitro Fertilization.



This article is an open access article distributed under the terms and conditions of the Creative Commons

Attribution (CC BY) license http://creativecommons.org/licenses/by/4.0/

Introduction

Melatonin is formed and released primarily by the pineal gland (1). Furthermore, it can be produced by other organs as it has been found in extra-pineal organs, including digestive system, brain, eye, respiratory system, skin, renal system, thyroid, thymus, immune system and reproductive system (2-12).

The discovery of melatonin by Lerner and team in 1958 has led to a new research area in the physiology of reproduction (1). Melatonin might have an essential role in regulating men and women fertility (2).

pineal organs as well. It's known as an organizer of circadian rhythms and more recently as an anti-oxidant. In addition to its role in maintaining immunity, pathophysiology of cardiovascular and neurological diseases, and as an anti-cancer agent. Evidence has demonstrated that melatonin exerts a positive impact on male and female fertility primarily through oxygen scavenging effects. Melatonin might have positive effects on nuclear and cytoplasmic maturation of oocytes and it probably opposes meiotic errors in old age oocytes. In Assisted reproductive technologies, supplementation of melatonin was shown be associated with better outcomes in terms of sperm quality, oocyte quality, embryo quality and pregnancy rates. Melatonin possibly shows promise as a supportive treatment in cases of infertility, thus trying to reach more understanding about its role in fertility is mandatory. Previous research has shown contradicting results regarding the role of melatonin in fertility. This review summarizes various actions of melatonin on the body focusing on male and female fecundity.

Melatonin, a hormone synthesized mainly by the pineal gland, has been found in extra-

Recently, it has been shown that oxidative stress, an imbalance between reactive oxygen and nitrogen species, and antioxidants, is a contributing factor for decreasing fertility in both genders (13, 14).

Evidence showed that oxidative stress might negatively influence the success of In vitro fertilization (IVF) programs (15), therefore, melatonin, as one of the novel oxygen scavengers, has been used to decrease oxidative stress and to enhance IVF outcome. Its antioxidant effects as a strong radical scavenger may enhance oocyte quality directly since it is a potent antioxidant present in the follicular fluid (2, 16, 17).

Melatonin probably acts as a supportive therapy in patients with infertility, thus trying to reach greater understanding about its mechanism of action in fecundity is essential.

This review aims to briefly discuss the impacts of melatonin on body systems shedding the light mainly on its role in male and female fertility.

Synthesis of Melatonin

Melatonin (N-acetyl-5-methoxytryptamine) is an Indoleamine that is produced from an essential amino acid called tryptophan. Its synthesis depends on the presence and absence of light as it is suppressed at daylight and stimulated at night (2). Thus, the release of melatonin displays a diurnal rhythm, started to be secreted in the evening, showing the highest level in the middle of the night (18).

Effects of melatonin on the body:

Melatonin has diverse range of effects on the body including its effects on the following systems:

Melatonin affects the immune system as Carrillo-Vico et al, suggested that melatonin can act as an immune buffer that boosts the immune system in basal and immunosuppressive states and suppresses the inflammation in conditions of exacerbation of immune reactions (19). The two types of melatonin receptors; the membrane and the nuclear receptors have been recognized on white blood cells. Membrane receptors were mainly present on CD4 T lymphocytes, CD8 T and B cells (20, 21). Melatonin regulates the production of cytokines by mononuclear cells (22). It has been stated that melatonin activates the innate immunity and the adaptive immunity. In contrast, melatonin can reduce the inflammation via the prevention of nuclear factor Kappa B binding to DNA and the inhibition of its translocation to the nucleus, these effects decrease cytokines and chemokines production (23).

Melatonin has been reported to be an anti-cancer agent as it acts against proliferation and metastasis, and it has pro-apoptotic and immunostimulatory properties (24), it has been suggested that it can be used in the treatment of some malignancies (24- 26). Melatonin may exert its anti- cancer action when it's used solely or combined with other anti- cancer therapies (27).

Evidence has proposed that melatonin has an essential role in the pathophysiology of some cardiovascular diseases (28, 29). A recent study has stated that melatonin can protect against myocardial damage resulted from high fat diet (30).

It has been shown that melatonin can maintain the integrity of blood brain barrier (31). Besides its ability of crossing the blood brain barrier, its capability to reduce oxidative stress and inflammation and its action as an anti- excitotoxicity and anti-misfolding make it a promising neuroprotector in some neurological diseases (32).

Melatonin and male fertility:

The hypothalamic pituitary testicular axis acts through positive and negative feedbacks, to regulate the function of the testis. The hypothalamus releases Gonadotropin releasing hormone (GnRH) in a pulsatile manner triggering the secretion of gonadotropins namely Follicle stimulating hormone (FSH) and Luteinizing hormone (LH) from the anterior pituitary gland, those hormones mediate testicular functions, steroidogenesis and spermatogenesis (33).

Melatonin has a role in regulating the release of GnRH and LH (34). In a study done on sheep, it has been shown that melatonin suppresses LH release (35). Another study done on neonatal rats showed that inhibiting LH secretion by melatonin could be explained by a decrease of both second messengers (Ca2+) and cyclic AMP (36).

However, no impact of melatonin was found on LH, FSH and testosterone levels when administered for long term to normal males (37). Thus, more research is mandatory to illustrate melatonin's influence on hypothalamic pituitary testicular axis.

Melatonin is considered as a regulator of testicular steroidogenesis (34). In mice that received melatonin treatment, there were reductions in nuclear volume, endoplasmic reticulum volume, mitochondria and Golgi complex volumes of mice Leydig cells suggesting that such effects can probably inhibit Leydig cells from secreting testosterone (38).

Melatonin administration can also decrease Leydig cell cyclic AMP concentrations which is important in testosterone synthesis via LH signaling and this effect can be abolished by giving luzindole which is a melatonin receptor antagonist that stimulates testicular steroidogenesis, melatonin also suppresses Leydig cell steroidogenesis through inhibiting the expression of Steroidogenic Acute Regulatory Protein in MA- 10 mice (39).

Spermatozoa are considered highly susceptible for oxidative stress (40). A recent study done on rabbit's spermatozoa revealed that melatonin protects spermatozoa from reactive oxygen species (ROS) by improving AMP activated protein kinase (AMPK) phosphorylation (41).

Regarding human spermatozoa, Deng et al, showed that addition of melatonin to the cryoprotectant during the process of semen cryopreservation improves spermatozoa viability, and membrane integrity, and reduces intracellular ROS and lipid peroxidation damage. Furthermore, melatonin enhances heat shock protein (HSP90) expression in frozen–thawed sperms through melatonin receptor MT1 (42). Another study has reported that melatonin has free radical scavenging effects because it opposed sperm apoptosis in ejaculated human spermatozoa (43).

A recent study done on bull spermatozoa suggested that melatonin has capacitation-modulating effect and protective action at physiological concentrations (44).

Various studies have mentioned a decline of semen quality all over the world (45- 48). Semen quality is considered a marker for male fertility, thus trials to find factors improving it are essential.

Researchers evaluated melatonin effects on the quality of rooster sperms comparing it with control samples with no added melatonin, found that sperm count is greater in samples with melatonin, cell membrane performance and mitochondrial function are also better in melatonin samples than the control samples. In addition, oxidative degradation of lipids, DNA fragmentation, and apoptosis were all less in samples treated with melatonin (49).

In human, post- thaw sperms that have been treated with various melatonin concentrations (0.001, 0.005, 0.01, 0.05, 0.1, and 1 mM), during the process of cryopreservation, showed better spontaneous

movement and survival with less intracellular ROS levels and malondialdehyde (MDA) levels than the control group in all melatonin doses except for 0.001 mM. Melatonin concentration of 0.01 mM was the most effective one in protecting spermatozoa from oxidative stress (50).

On the other hand, it was shown that administration of melatonin can negatively affect the semen quality in healthy men, this could be due to aromatase inhibition (51). However, in this study long term administration of melatonin was used and the study was done on healthy men.

Sharbatoghli et al, have not found a relationship between seminal plasma melatonin and sperm parameters, nonetheless, they showed that DNA fragmentation and melatonin levels are positively associated in infertile men (52).

A study done on human semen samples showed that melatonin reduces ROS that are derived from the mitochondria and it rescues the decreased penetration ability of spermatozoa treated with 3-Nitrophthalic acid (3-NPA) thus melatonin might have the clinical potential to enhance sperm quality (53).

A recent research compared the seminal plasma and serum melatonin levels of men with idiopathic oligoasthenoteratozoospermia and normal, fertile males. It evaluated the effects of exposure to light at night on semen parameters, the study demonstrated a significantly lower serum and seminal plasma levels of melatonin in patients exposed to light at night in comparison to non-exposed cases. Therefore, darkness and sleep at night-time may potentiate the semen parameters of males with idiopathic oligoasthenoteratozoospermia (54).

So, adequate melatonin level is crucial for improving male fertility. Its benefit is possibly evident mainly in cases of idiopathic infertility, however, administration of melatonin for long duration is not advised and further studies are necessary in this research area to illustrate the mechanisms of melatonin impact on semen quality.

Melatonin and female fertility:

Oxidative stress exerts toxic effects on the oocytes maturation. Evidence showed that melatonin might protect oocytes from free radical damage (55).

The process of oxidative damage in mice oocytes may happen as soon as eight hours following being in the culture, and it coexists with the appearance of markers of apoptosis like phosphatidylserine externalization, then, after 16 hours, caspase activation takes place along with structural alterations of oocyte biological aging as shown by Lord et al (56). The researchers of the same study also concluded that melatonin supplementation can prevent oxidative stress hence delay the aging process of mice oocytes, so it might be useful in clinical practice (56).

A review stated that ROS are produced in the follicles mainly during ovulation, they can be scavenged by melatonin thus reducing oxidative stress and improving oocyte maturation as illustrated in figure 1 (57).

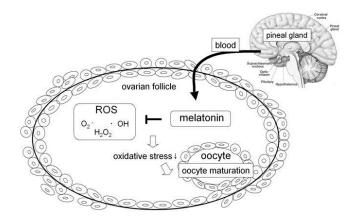


Figure 1. Role of melatonin as an antioxidant in ovarian antral follicle (57). This figure was published by BioMed Central Ltd/ part of Springer Nature

(https://ovarianresearch.biomedcentral.com/articles/10.1186/1757-

2215-5-5/figures/4), distributed under the terms of the Creative Commons Attribution License 2.0

(http://creativecommons.org/licenses/by/2.0) *the title of this figure is modified from the original reference.

In a study done by Kang et al, the researchers compared porcine oocytes treated with melatonin and those that are not treated, during the process of In vitro maturation (IVM). It was found that ROS are significantly lower in oocytes treated with melatonin. Also, melatonin has had positive effects on nuclear and cytoplasmic maturation of porcine oocytes (58). Another study has demonstrated that melatonin improves oocyte maturation and thus it possibly enhances oocyte quality and fertilization rates (55).

Evidence showed that melatonin administration potentiates oocyte maturation, embryo cleavage and blastocyst development rates in mice, nonetheless, the response to melatonin differs according to the doses used in various stages of maturation and development since cumulus oocyte complexes required lower melatonin concentration during maturation stages compared with oocytes without cumulus cells and embryos needed lower melatonin concentrations in this experiment (59).

Regarding human oocytes, Wei and coworkers, evaluated nuclear maturation of oocytes that were cultured in mediums treated with different melatonin concentrations, they found that low melatonin concentrations enhances nuclear maturation, whereas, high concentrations of melatonin decreases nuclear maturation (60). Further research is probably needed in this field to reach more understanding.

Yang et al, illustrated that melatonin can prevent oocyte aging following ovulation in mice as it decreased ROS and inhibited meiosis abnormalities, mitochondrial disorders, autophagocytosis and apoptosis via the upregulation of SIRT1 and MnSOD protein levels in postovulatory aged oocytes (61).

A more recent study, done in 2020, found that melatonin in the follicular fluid reduces with age in mice. Supplementation of old mice with melatonin can protect oocytes from spindle and chromosomal abnormalities and from aneuploidies. Melatonin opposes meiotic errors in old age oocytes via stimulation of Sirt1/Sod2 pathway. The increment of Sirt1 expression in oocytes treated with melatonin promoted the expression of Sod2 thus reducing ROS and improving oocyte quality (62).

In women with Polycystic ovary syndrome (PCOS), it has been suggested that melatonin enhances the expression of cytochrome P450 family 19 subfamily A member 1 (CYP19A1) and heme oxygenase-1 (HO-1), and decreases the levels of interleukin 18 (IL-18) in the granulosa cells thus improving oocyte maturation in PCOS women with high androgen levels (63).

A research showed that women with unexplained infertility can also benefit from melatonin administration as two doses (three mg/day or six mg/day) of melatonin have been tried which probably led to decrement of oxidative stress and improvement of oocyte quality in patients with idiopathic infertility. Nonetheless, the authors of the study have encouraged more studies in people with various backgrounds to confirm the importance of melatonin in treating infertility (64).

Systematic review and meta-analysis of randomized controlled trials concluded that more research, regarding melatonin role in treating infertility, is needed before the application of the routine use of melatonin in practice (65).

Melatonin and In Vitro Fertilization (IVF):

In order to get high quantity of oocytes and to achieve good pregnancy rates, ovarian stimulation protocols are used in IVF (66). Nevertheless, they can increase oxidative stress since they might alter the follicular environment and affect the endogenous levels of oxygen scavengers (2, 66). Follicular fluid has strong anti-oxidant activity, however, the in vitro oocytes are not protected by this fluid thus they are highly susceptible to oxidative stress (2, 67). In addition, the high oxygen concentrations that oocytes may expose to in the incubators and throughout dealing with them in IVF procedures may increase ROS (2).

Reactive oxygen species generation in granulosa cells of females having IVF, mainly in cases of PCOS, might affect IVF success rates negatively (68). It has been suggested that the administration of micronutrients may exert positive effects on IVF results possibly by decreasing oxidative damage in serum and follicular fluid proteins thus improving oocyte quality (69).

A study demonstrated that treating women, who have unsuccessful IVF cycles previously, with melatonin can improve oocyte quality since the administration of melatonin tablet (three mg) can increase the concentration of intrafollicular melatonin and it can reduce the concentration of intrafollicular lipid peroxide (70). Rizzo et al, found that women with history of low oocyte quality who were given melatonin in addition to myo-inositol and folic acid in IVF cycles have had better oocyte quality, and pregnancy outcome than women who received myo-inositol and folic acid without melatonin (17).

In addition to the influence of melatonin on the sperms and the oocytes as mentioned previously, studies have found that it has beneficial effects on the embryos as well (71-73). The environment that the embryos are cultured in is important to determine the success of fertilization and implantation (2). Rodriguez- Osorio and team found a positive impact of melatonin on porcine embryo

cleavage rate and blastocyst cell numbers at a concentration of 10-9 m (71). In mice, it was found that melatonin potentiates the early growth of the embryos and the development rate to blastocysts, probably through its effect on the metabolism in early embryogenesis stages thus stimulating blastocysts formation (72). Also, melatonin was found to have beneficial influences on the embryos of the sheep through two mechanisms, by reducing the oxidative stress and by affecting the fertilization process positively (73).

Luteal phase insufficiency can result from free radical damage which might reduce the progesterone levels (74). Melatonin may support the luteal phase as a study showed that it potentiates progesterone production in human granulosa lutein cells (75). Another study, done on 66 women, showed that melatonin increases progesterone levels significantly in the treatment cycle in comparison to the preceding one, however, in the same study, it was noticed that melatonin enhances progesterone concentration nonsignificantly in the patients in comparison to the controls (76). Thus, more research is needed to investigate the effects of melatonin in supporting the luteal phase.

Studies have shown that melatonin might improve pregnancy rates (17, 55, 77). In a prospective trial involved 65 women, Rizzo et al, found that women with history of low oocyte quality who were given melatonin in addition to myo-inositol and folic acid in IVF cycles have had in tendency better clinical pregnancy rate compared with women who were not received melatonin, though the variation was not significant (17). So, using a combination of antioxidants during the treatment of infertility might improve the outcome. Tamura et al, have observed that the gestation rate was higher in patients received melatonin than patients with no melatonin (55). A randomized double blinded clinical trial was performed on PCOS patients underwent intrauterine insemination (IUI) treatment, compared two groups in term of chemical pregnancy rates; the group of women who were given melatonin, and the control group, the study showed that chemical pregnancy rates were 32% and 18% respectively with a p value of 0.012 (77).

Melatonin can be used as a supportive therapy in treating male infertility mainly in cases of unexplained infertility and in treating female infertility, especially in PCOS and unexplained infertility conditions. Studies showed contradicting results which could be due to heterogeneity of the patients involved in the studies, in terms of reasons of infertility, age groups and doses of melatonin used. Thus, more randomized controlled trials are essential to be done taking into consideration causes of these controversial findings.

Conclusion

Melatonin has been known as an anti-oxidant that exerts various effects on the body. It has also been used as an adjunctive treatment in cases of male and female infertility to reduce oxidative stress that can be associated with infertility, thus improving success rates of pregnancy mainly in IVF programs. However, its routine use in practice still needs further well-designed studies such as more randomized controlled trials in the field of fertility.

Funding

This research did not receive any specific fund.

Conflict of Interest

No conflict of interest

References

- Lerner AB, Case JD, Takahashi Y, Lee TH, Mori WJJotACS. Isolation of melatonin, the pineal gland factor that lightens melanocyteS1. 1958;80(10):2587.-
- [2] Fernando S, Rombauts LJJoor. Melatonin: shedding light on infertility?-a review of the recent literature. 2014;7(1):1-14.
- [3] Madalinski MHJWjogp, therapeutics. Does a melatonin supplement alter the course of gastro-esophageal reflux disease? 2011;2(6):50.
- [4] Carbajo-Pescador S, Ordoñez R, Benet M, Jover R, García-Palomo A, Mauriz J, et al. Inhibition of VEGF expression through blockade of Hifl α and STAT3 signalling mediates the anti-angiogenic effect of melatonin in HepG2 liver cancer cells. 2013;109(1):83-91.
- [5] Jaworek J, Leja-Szpak A, Nawrot-Porąbka K, Szklarczyk J, Kot M, Pierzchalski P, et al. Effects of melatonin and its analogues on pancreatic inflammation, enzyme secretion, and tumorigenesis. 2017;18(5):1014.
- [6] Lin YW, Chen TY, Hung CY, Tai SH, Huang SY, Chang CC, et al. Melatonin protects brain against ischemia/reperfusion injury by attenuating endoplasmic reticulum stress. 2018;42(1):182-92.
- [7] Blasiak J, Reiter RJ, Kaarniranta KJOm, longevity c. Melatonin in retinal physiology and pathology: the case of age-related macular degeneration. 2016;2016.
- [8] de Matos Cavalcante AG, de Bruin PFC, de Bruin VMS, Nunes DM, Pereira EDB, Cavalcante MM, et al. Melatonin reduces lung oxidative stress in patients with chronic obstructive pulmonary disease: a randomized, double- blind, placebo- controlled study. 2012;53(3):238-44.
- [9] Scheuer C, Pommergaard HC, Rosenberg J, Gögenur IJP, photoimmunology, photomedicine. Melatonin's protective effect against UV radiation: a systematic review of clinical and experimental studies. 2014;30(4):180-8.
- [10] Khodadadi S, Nasri P, Ardalan M-R, Rafieian-Kopaei MJAoRiA. Melatonin and kidney; a narrative review on the renoprotective efficacy of melatonin in various renal diseases. 2016;1.(^Y)
- [11] Csaba GJAmeiH. The pineal regulation of the immune system: 40 years since the discovery. 2013;60(2):77-91.

- [12] Hardeland RJBJSTR. Melatonin—More than just a pineal hormone. 2017;1(4):1-4.
- [13] Agarwal A, Makker K, Sharma RJAjori. Clinical relevance of oxidative stress in male factor infertility: an update. 2008;59(1):2-11.
- [14] Agarwal A, Aponte-Mellado A, Premkumar BJ, Shaman A, Gupta SJRb, endocrinology. The effects of oxidative stress on female reproduction: a review. 2012;10(1):1-31.
- [15] Sinha A, Gupta S. Oxidative Stress in Assisted Reproductive Technology. 2016.
- [16] Tong J, Sheng S, Sun Y, Li H, Li W-P, Zhang C, et al. Melatonin levels in follicular fluid as markers for IVF outcomes and predicting ovarian reserve. 2017;153(4):443-51.
- [17] Rizzo P, Raffone E, Benedetto VJERMPS. Effect of the treatment with myo-inositol plus folic acid plus melatonin in comparison with a treatment with myoinositol plus folic acid on oocyte quality and pregnancy outcome in IVF cycles. A prospective, clinical trial. 2010;14(6):555-61.
- [18] Liu S, Madu C, Lu Y. The Role of Melatonin in Cancer Development. Oncomedicine 2018; 3: 37-47. doi: 10.7150/oncm. 25566.
- [19] Carrillo-Vico A, Lardone PJ, Álvarez-Sánchez N, Rodríguez-Rodríguez A, Guerrero JMJIjoms. Melatonin: buffering the immune system. 2013;14(4):8638-83.
- [20] Emet M, Ozcan H, Ozel L, Yayla M, Halici Z, Hacimuftuoglu AJTEjom. A review of melatonin, its receptors and drugs. 2016;48(2):135.
- [21] Pozo D, Delgado M, Fernandez- Santos JM, Calvo JR, Gomariz RP, Martin- Lacave I, et al. Expression of the Mel1a- melatonin receptor mRNA in T and B subsets of lymphocytes from rat thymus and spleen. 1997;11(6):466-73.
- [22] García-Mauriño S, Pozo D, Carrillo-Vico A, Calvo JR, Guerrero JMJLs. Melatonin activates Th1 lymphocytes by increasing IL-12 production. 1999;65(20):2143-50.
- [23] Szczepanik MJJop, pharmacology. Melatonin and its influence on immune system. 2007;58(6):115-24.
- [24] Ma Z, Yang Y, Fan C, Han J, Wang D, Di S, et al. Melatonin as a potential anticarcinogen for non-smallcell lung cancer. 2016;7(29):46768.
- [25] Yang C-Y, Lin C-K, Tsao C-H, Hsieh C-C, Lin G-J, Ma K-H, et al. Melatonin exerts anti-oral cancer effect via suppressing LSD1 in patient-derived tumor xenograft models. 2017;8(20):33756.
- [26] Song J, Ma S-J, Luo J-H, Zhang H, Wang R-X, Liu H, et al. Melatonin induces the apoptosis and inhibits the

proliferation of human gastric cancer cells via blockade of the AKT/MDM2 pathway. 2018;39(4):1975-83.

- [27] Talib WHJM. Melatonin and cancer hallmarks. 2018;23(3):518.
- [28] Tengattini S, Reiter RJ, Tan DX, Terron MP, Rodella LF, Rezzani RJJopr. Cardiovascular diseases: protective effects of melatonin. 2008;44(1):16-25.
- [29] Nduhirabandi F, Maarman GJJM. Melatonin in heart failure: a promising therapeutic strategy? 2018;23(7):1819.
- [30] Bose G, Ghosh A, Chattopadhyay A, Pal PK, Bandyopadhyay DJMR. Melatonin as a potential therapeutic molecule against myocardial damage caused by high fat diet (HFD). 2019;2(3):37-56.
- [31] Liu W-C, Wang X, Zhang X, Chen X, Jin XJFian. Melatonin supplementation, a strategy to prevent neurological diseases through maintaining integrity of blood brain barrier in old people. 2017;9:165.
- [32] Alghamdi BJJonr. The neuroprotective role of melatonin in neurological disorders. 2018;96(7):1136-49.
- [33] Dutta S, Sengupta P, Muhamad SJAPJoR. Male reproductive hormones and semen quality. 2019;8(5):189.
- [34] Bhattacharya K, Sengupta P, Dutta SJAPJoR. Role of melatonin in male reproduction. 2019;8(5):211.
- [35] Misztal T, Romanowicz K, Barcikowski BJRB. Melatonin-a modulator of the GnRH/LH axis in sheep. 2002;2(3):267-75.
- [36] Vaneček JJPR. Decrease of [Ca**], and Cyclic AMP. 1998;47:329-35.
- [37] Luboshitzky R, Levi M, Shen-Orr Z, Blumenfeld Z, Herer P, Lavie PJHr. Long-term melatonin administration does not alter pituitary-gonadal hormone secretion in normal men. 2000;15(1):60-5.
- [38] Redins C, Redins G, Novaes JJBJoB. The effects of treatment with melatonin on the ultrastructure of mouse Leydig cells: a quantitative study. 2002;62(3):517-23.
- [39] WU CS, LEU SF, YANG HY, HUANG BMJJoa. Melatonin inhibits the expression of steroidogenic acute regulatory protein and steroidogenesis in MA- 10 cells. 2001;22(2):245-54.
- [40] Bennetts LE, Aitken RJJMR, Research DIG. A comparative study of oxidative DNA damage in mammalian spermatozoa. 2005;71(1):77-87.
- [41] Zhu Z, Li R, Lv Y, Zeng WJC. Melatonin protects rabbit spermatozoa from cryo-damage via decreasing oxidative stress. 2019;88:1-8.
- [42] Deng S-L, Sun T-C, Yu K, Wang Z-P, Zhang B-L, Zhang Y, et al. Melatonin reduces oxidative damage and

upregulates heat shock protein 90 expression in cryopreserved human semen. 2017;113:347-54.

- [43] Espino J, Bejarano I, Ortiz Á, Lozano GM, García JF, Pariente JA, et al. Melatonin as a potential tool against oxidative damage and apoptosis in ejaculated human spermatozoa. 2010;94(5):1915-7.
- [44] Fernández-Alegre E, Álvarez-Fernández I, Domínguez JC, Casao A, Martínez-Pastor FJIjoms. Melatonin nonlinearly modulates bull spermatozoa motility and physiology in capacitating and non-capacitating conditions. 2020;21(8):2701.
- [45] Huang C, Li B, Xu K, Liu D, Hu J, Yang Y, et al. Decline in semen quality among 30,636 young Chinese men from 2001 to 2015. 2017;107(1):83-8. e2.
- [46] Borges E, Setti AS, Braga DPdAF, Figueira RdCS, Iaconelli AJIbju. Decline in semen quality among infertile men in Brazil during the past 10 years. 2015;41:757-63.
- [47] Li C-J, Tzeng C-R, Chen R-Y, Han B-C, Yeh C-Y, Chien L-CJCJP. Decline in semen quality in men in northern Taiwan between 2001 and 2010. 2016;59(6):355-65.
- [48] Geoffroy-Siraudin C, Anderson Dieudonné Loundou FR, Achard V, Courbiere B, Perrard M-H, Durand P, et al. Decline of semen quality among 10 932 males consulting for couple infertility over a 20-year period in Marseille, France. 2012;14(4):584.
- [49] Mehaisen GM, Partyka A, Ligocka Z, Niżański WJArs. Cryoprotective effect of melatonin supplementation on post-thawed rooster sperm quality. 2020;212:106238.
- [50] Karimfar M, Niazvand F, Haghani K, Ghafourian S, Shirazi R, Bakhtiyari SJIjoi, et al. The protective effects of melatonin against cryopreservation-induced oxidative stress in human sperm. 2015;28(1):69-76.
- [51] Luboshitzky R, SHEN- ORR Z, Nave R, Lavi S, Lavie PJJoa. Melatonin administration alters semen quality in healthy men. 2002;23(4):572-8.
- [52] Sharbatoghli M, Valojerdi MR, Bahadori MH, Yazdi RS, Ghaleno LRJCJ. The relationship between seminal melatonin with sperm parameters, DNA fragmentation and nuclear maturity in intra-cytoplasmic sperm injection candidates. 2015;17(3):547.
- [53] Zhang X-Y, Xiong Y-M, Tan Y-J, Wang L, Li R, Zhang Y, et al. Melatonin rescues impaired penetration ability of human spermatozoa induced by mitochondrial dysfunction. 2019;158(5):465-75.
- [54] Hassan MH, El- Taieb MA, Fares NN, Fayed HM, Toghan R, Ibrahim HMJE, et al. Men with idiopathic oligoasthenoteratozoospermia exhibit lower serum and seminal plasma melatonin levels: Comparative effect of

night-light exposure with fertile males. 2020;20(1):235-42.

- [55] Tamura H, Takasaki A, Miwa I, Taniguchi K, Maekawa R, Asada H, et al. Oxidative stress impairs oocyte quality and melatonin protects oocytes from free radical damage and improves fertilization rate. 2008;44(3):280-7.
- [56] Lord T, Nixon B, Jones KT, Aitken RJJBor. Melatonin prevents postovulatory oocyte aging in the mouse and extends the window for optimal fertilization in vitro. 2013;88(3):67, 1-9.
- [57] Tamura H, Takasaki A, Taketani T, Tanabe M, Kizuka F, Lee L, et al. The role of melatonin as an antioxidant in the follicle. 2012;5(1):1-9.
- [58] Kang JT, Koo OJ, Kwon DK, Park HJ, Jang G, Kang SK, et al. Effects of melatonin on in vitro maturation of porcine oocyte and expression of melatonin receptor RNA in cumulus and granulosa cells. 2009;46(1):22-8.
- [59] Bahadori MH, Ghasemian F, Ramezani M, Asgari ZJIjorm. Melatonin effect during different maturation stages of oocyte and subsequent embryo development in mice. 2013;11(1):11.
- [60] Wei D, Zhang C, Xie J, Song X, Yin B, Liu Q, et al. Supplementation with low concentrations of melatonin improves nuclear maturation of human oocytes in vitro. 2013;30(7):933-8.
- [61] Yang Q, Dai S, Luo X, Zhu J, Li F, Liu J, et al. Melatonin attenuates postovulatory oocyte dysfunction by regulating SIRT1 expression. 2018;156(1):81-92.
- [62] Zhang M, Lu Y, Chen Y, Zhang Y, Xiong BJRb. Insufficiency of melatonin in follicular fluid is a reversible cause for advanced maternal age-related aneuploidy in oocytes. 2020;28:101327.
- [63] Yu K, Wang R-X, Li M-H, Sun T-C, Zhou Y-W, Li Y-Y, et al. Melatonin reduces androgen production and upregulates heme oxygenase-1 expression in granulosa cells from PCOS patients with hypoestrogenia and hyperandrogenia. 2019;2019.
- [64] Espino J, Macedo M, Lozano G, Ortiz Á, Rodríguez C, Rodríguez AB, et al. Impact of melatonin supplementation in women with unexplained infertility undergoing fertility treatment. 2019;8(9):338.
- [65] Seko LM, Moroni RM, Leitao VM, Teixeira DM, Nastri CO, Martins WPJF, et al. Melatonin supplementation during controlled ovarian stimulation for women undergoing assisted reproductive technology: systematic review and meta-analysis of randomized controlled trials. 2014;101(1):154-61. e4.
- [66] Palini S, Benedetti S, Tagliamonte MC, De Stefani S, Primiterra M, Polli V, et al. Influence of ovarian

stimulation for IVF/ICSI on the antioxidant defence system and relationship to outcome. 2014;29(1):65-71.

- [67] Huang B, Li Z, Ai J, Zhu L, Li Y, Jin L, et al. Antioxidant capacity of follicular fluid from patients undergoing in vitro fertilization. 2014;7(5):2273.
- [68] Karuputhula NB, Chattopadhyay R, Chakravarty B, Chaudhury KJSbirm. Oxidative status in granulosa cells of infertile women undergoing IVF. 2013;59(2):91-8.
- [69] Luddi A, Capaldo A, Focarelli R, Gori M, Morgante G, Piomboni P, et al. Antioxidants reduce oxidative stress in follicular fluid of aged women undergoing IVF. 2016;14(1):1-7.
- [70] Takasaki A, Nakamura Y, Tamura H, Shimamura K, Morioka HJRm, biology. Melatonin as a new drug for improving oocyte quality. 2003;2(4):139-44.
- [71] Rodriguez- Osorio N, Kim I, Wang H, Kaya A, Memili EJJopr. Melatonin increases cleavage rate of porcine preimplantation embryos in vitro. 2007;43(3):283-8.
- [72] Ishizuka B, Kuribayashi Y, Murai K, Amemiya A, Itoh MTJJopr. The effect of melatonin on in vitro fertilization and embryo development in mice. 2000;28(1):48-51.
- [73] Abecia J-A, Forcada F, Vázquez M-I, Muiño-Blanco T, Cebrián-Pérez JA, Pérez-Pe R, et al. Role of melatonin on embryo viability in sheep. 2019;31(1):82-92.
- [74] Shi L, Zhang J, Lai Z, Tian Y, Fang L, Wu M, et al. Long-term moderate oxidative stress decreased ovarian reproductive function by reducing follicle quality and progesterone production. 2016;11(9):e0162194.
- [75] Fang L, Li Y, Wang S, Yu Y, Li Y, Guo Y, et al. Melatonin induces progesterone production in human granulosa-lutein cells through upregulation of StAR expression. 2019;11(20):9013.
- [76] Takasaki A, Tamura H, Taniguchi K, Asada H, Taketani T, Matsuoka A, et al. Luteal blood flow and luteal function. 2009;2(1):1-6.
- [77] Mokhtari F, Asbagh FA, Azmoodeh O, Bakhtiyari M, Almasi-Hashiani AJIjof, sterility. Effects of melatonin administration on chemical pregnancy rates of polycystic ovary syndrome patients undergoing intrauterine insemination: a randomized clinical trial. 2019;13(3):225.

To cite this article: Alawad Z, Al-Omary H. Melatonin in male and female fertility. Al-Kindy College Medical Journal. 2021;17(3):145-151.