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Preparation of Vaccine against Diabetic Foot Pathogenic Bacteria Using Low Level Diode Laser

ARTICLE INFORMATION

ABSTRACT

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Background: Since the invention of laser in 1960, lasers have been developed and approved in many fields. Lasers can now be regarded as practical tools with unique properties that have been utilized effectively in several applications in fields of medical and biological sciences.

Objectives: The aim of the current study was to preparation of vaccines (live attenuated and killed) by irradiation of the bacteria by the low level diode laser.

Methods: six bacterial isolates were isolated from human samples of diabetic foot infections, which used for preparation of vaccines. The experiment was conducted on fifteen adult male rabbits; they were divided into three groups with 5 rabbits each. Blood samples were collected from the marginal ear vein of the rabbits after one month of the vaccination, for the purpose of measuring the concentration of the immunoglobulins which present in their serum, using Radial Immunodiffusion (RID) method by specialized kits (LTA-Italy).

Results: After irradiation of the bacterial suspensions with the diode laser for different exposure times and different frequencies, and the wavelengths used were (660, 820, 915) nm, the growth of bacterial isolates decreased until killed of bacteria at 40 min. The results of IgA concentrations for the three groups were highly significant ($P < 0.01$) when comparing the attenuated with control group, while were significant ($P < 0.05$) between the killed vaccine and control groups, and not significant variations between the attenuated and killed groups,

Conclusions: Wavelength (660) nm was more effective in killing the bacteria, and the variations were not significant between the live attenuated and the killed vaccine.

Introduction:

LASER is an acronym for light amplification by stimulated emission of radiation; common usage today is to use the word as a noun (laser) rather than as an acronym⁽¹⁾. Laser is a device that converts electrical or chemical energy into light energy. In contrast to ordinary light that is emitted spontaneously by excited atoms or molecules, the light emitted by laser occurs when an atom or molecule retains excess energy until it is stimulated to emit it. The radiation emitted by lasers including both visible and invisible light is more generally termed as electromagnetic radiation⁽²⁾.

Diabetic Foot Ulcers (DFUs) are a common and much feared complication of diabetes, with recent studies suggesting that the lifetime risk of developing a foot ulcer in diabetic patients may be as high as 25%⁽³⁾. Infection is a frequent (40%-80%) and costly complication of these ulcers and represents a major cause of morbidity and mortality. It is estimated to be the most common cause of diabetes-related admission to hospital and remains one of the major pathways to lower-limb amputation⁽⁴⁾.

A vaccine is a suspension that contains a part of a pathogen that induces the immune system to produce antibodies that combat the antigen⁽⁵⁾. When the vaccine is

injected into a body the chemicals in the vaccine cause tissue irritation, this result in blood flow to the injection site and with the blood come white blood cells, the white blood cells become exposed to the antigen and begin a series of processes that cause antibodies to be produced to the antigen. The period of time from when the vaccine is injected until production of antibodies takes days or weeks⁽⁶⁾.

The immunoglobulins are Proteins, present on the surface of B lymphocytes, secreted in response to stimulation, that neutralize antigens by binding specifically to their surface⁽⁷⁾.

Methods:

Bacterial species: six bacterial isolates (Staphylococcus aureus, S. epidermidis, Pseudomonas aeruginosa, Escherichia coli, Proteus mirabilis, and Klebsiella pneumonia) were isolated from human samples which collected from patients admitted to Al- Hussain Teaching Hospital, Al- Muthanna Province, suffering from diabetic foot infections, using sterilized cotton swabs. These samples were identified according to Bergeys manual using different morphological and biochemical tests⁽⁸⁾.

Vaccine preparation: six isolates of bacteria were cultured on blood agar at 37°C for 24hr. washing the

surface of the plates with normal saline using glass rods. The bacterial suspension mixed with vortex for 3 min. five milliliters of the suspension from each culture was cold centrifugation at 6000 r.p.m for 10 min. The sediment of the bacteria washed three times by normal saline (pH=7.2) then mixed by vortex to be suspended once again in 5 ml of normal saline and compared with Macferland solution. The bacterial suspension irradiated with laser to obtain live attenuated and killed vaccines⁽⁹⁾. Then bacterial suspension recultured on blood agar and incubated at 37°C for 24h to determine the live attenuated and killed bacteria in which the growth not observed.

Vaccine program: Fifteen male rabbits were divided into three groups and injected as following; First group: n =5 inoculated with live attenuated vaccine (A vaccine) consist of a mixture of (*S. aureus*, *S. epidermidis*, *Pseudomonas aeruginosa*, *Escherichia coli*, *Proteus mirabilis*, and *Klebsiella pneumonia*) 1.8ml (0.3ml from each one) subcutaneously. Second group: n =5 inoculated with killed vaccine (K vaccine) consist of a mixture of bacterial isolates 1.8ml (0.3ml from each one) subcutaneously. Third group: n=5 (control group) injected with physiological normal saline.

Collection of Blood samples: 3 ml blood samples were collected from the marginal ear vein of the rabbits after one month of the vaccination, for the purpose of measuring the concentration of the immunoglobulins which present in their serum. By using Radial Immunodiffusion (RID) method, using specialized kits (LTA-Italy).

Statistical Analysis: all the results obtained were estimated statistically by using Minitab program 2 sample T - test.

Results:

Radial Immunodiffusion (RID) method used for determination immunoglobulins concentration (IgG, IgA, IgM, C₃, and C₄), present in the serum of the rabbits, which were inoculated with live attenuated and killed vaccine. There was an increasing in the level of IgG for all the animals groups. The variations between the live attenuated vaccine group and the control group were very highly significant (P < 0.001), but were high significant (P < 0.01) between the killed vaccine group and the control one, and not significant between the attenuated vaccine group and the killed vaccine one, as observed in Table 1.

Table1: Mean Immunoglobulins concentration of the live attenuated vaccine , killed vaccine and control groups.

Immunoglobulins concentration	Groups		
	Attenuated	Killed	Control
IgG (mg/dL)	2750 ***	2335 **	1656
IgA (mg/dL)	906 **	820.9 *	551
IgM (mg/dL)	363.0 *	313.8 *	223.2
C ₃ (mg/dL)	300.0 *	299.9 *	234.5
C ₄ (mg/dL)	100.44 *	97.00 *	75.4

(***) Very high significant (P < 0.001); (**) Highly significant (P < 0.01); (*) Significant (P < 0.05).

The results of IgA concentrations for the three groups were highly significant (P < 0.01) when comparing the attenuated with control group, while were significant (P < 0.05) between the killed vaccine and control groups, and

not significant variations between the attenuated and killed groups, as shown in Table 1.

The same results were obtained for IgM, C₃ and C₄ concentrations, which showed significant variations (P < 0.05), for the both groups, attenuated and killed when compared with the control group, but not significant between the attenuated and killed groups, as shown in Table 1.

Discussion:

The concentrations of IgG showed very high significant variations, these results agreed with Brown et.al⁽¹⁰⁾, who found that IgG responses were significantly greater in the mice groups immunized with both subunits, 10 µg of antigen (*S. aureus*) mixed with 3 µg of cholera toxin. While the results of IgA concentrations were highly significant when comparing the attenuated vaccine with control groups, these results nearly agreed with Eijgenraam et.al⁽¹¹⁾, who measured the level of IgA specific for cholera toxin subunit B in serum before and after immunization. There were significant increase in IgA (P < 0.001), in both the control and the patient groups.

The concentrations of IgM for the three groups showed significant variations, these results not agreed with Ostensson and Lun⁽¹²⁾, who found after *E.coli* endotoxin infusion the Ig concentrations increased rapidly with significantly higher (IgM p < 0.001, and IgA p < 0.05), and also not agreed with the results of Russo et.al⁽¹³⁾, who reported that immunization with a formalin-killed *E.coli* strain gives significant results in serum IgG and IgA response directed against the immunizing strain that is comparable to that observed after immunization with a live strain. The main role of secretory IgA is to inhibit bacterial attachment and neutralize viruses in mucosal tissue. In addition IgA, but not IgG, are translocated across epithelial tissue and can neutralize viruses intracellularly. This indicates that IgA is the first line of defense in the mucosal compartment. Secretory IgA is generally considered to be a non inflammatory antibody because it does not trigger inflammatory processes when it binds to antigens⁽¹⁴⁾.

The results of complement C₃ and C₄ concentrations for the three groups showed significant variations (p < 0.05, for the both groups, attenuated, and killed) when compared with the control group, these results not agreed with those got by Bogaert et.al⁽¹⁵⁾, who recalled that the variations were not significant in colonization density between non-bacteremic and bacteremic complement-depleted mice or between bacteremic complement-depleted and control mice. Complement is central to innate humoral immunity, interacting with a host of soluble and membrane proteins, in addition to the anti-bacterial activity of the complement cascade⁽¹⁶⁾.

Conclusions:

Wavelength (660) nm was more effective in killing the bacteria, and the variations were not significant between the live attenuated and the killed vaccine.

References:

1. Issa, MM. The Evolution of Laser Therapy in the Treatment of Benign Prostatic Hyperplasia. Med. Reviews, LLC. 2005; 7(9): 15 - 22.

2. Jyoti N, Pankaj M, Tulika G and Shelly A. Dental Laser - A boon to prosthodontics. *International Journal of dental clinics*. 2010; 2 (2): 13- 21.
3. Prompers L, Schaper N, Apelqvist J, Edmonds M, Mauricio D, Uccioli L, Bakker K, Holstein P, Jirkovska A, Piaggese A, Reike H, Spraul M and Huijberts M. Prediction of outcome in individuals with diabetic foot ulcers: focus on the differences between individuals with and without peripheral arterial disease. *Diabetologia* 2008; 51:747-755.
4. Richard JL, Sotto A and Lavign JP. New insights in diabetic foot infection. *World J. Diabetes* 2011; 2(2): 24-32.
5. Betsy T and Keogh J. (2005). *Vaccines and Diagnosing Diseases: Microbiology demystified*. chap. (15). The McGraw-Hill Companies, Inc. pp: 223 - 245.
6. Goelz JL. *Basic Immunology*. *International Sheep Letter*. Pipestone Veterinary Clinic (2000); 20 (4).
7. Pashine A, Valiante NM and Ulmer JB. Targeting the innate immune response with improved vaccine adjuvants. *Nat Med* 2005; 11(5):63-68.
8. Monica C. (2006). *Microbiological tests: District Laboratory Practice in Tropical Countries*. 2nd ed. Chap. (2). Published in the United States of America by Cambridge University Press, New York. pp: 62 - 70.
9. Beaglehole R, Bonita R and Kjellstrom T. (2000). *Basic epidemiology*. WHO Library cataloguing - publication Data.
10. Brown E, Dumitrescu O, Thomas D, Badiou C, Koers M, Choudhury P, Vazquez V, Etienne J, Lina G, Vandenesch F and Bowden M. The Panton-Valentine leukocidin vaccine protects mice against lung and skin infections caused by *Staphylococcus aureus* USA300. *Clin. Microbiol. Infect.* 2009; 15(2): 156-164.
11. Eijgenraam W, Oortwijn B, Kamerling S, Fijter J, Bake A, Daha M and Kooten C. Secretory immunoglobulin A (IgA) responses in IgA nephropathy patients after mucosal immunization, as part of a polymeric IgA response. *Clinical and Experimental Immunology* 2008; 152: 227-232.
12. Ostensson K and Lun L. Transfer of immunoglobulins through the mammary endothelium and epithelium and in the local lymph node of cows during the initial response after intramammary challenge with *E. coli* endotoxin. *Acta Veterinaria Scandinavica* 2008; 50 (26).
13. Russo T, Beanan M, Olson R, Genagon S, MacDonald U, Cope J, Davidson B, Johnston B and James R. A Killed, Genetically Engineered Derivative of a Wild-Type Extraintestinal Pathogenic *E. coli* strain is a Vaccine Candidate. *Vaccine* 2007; 25(19): 3859-3870.
14. Olas H, Butterweck W, Teschner H, Schwarz P and Reipert N. Immuno modulatory properties of human serum immunoglobulin A: anti-inflammatory and pro inflammatory activities in human monocytes and peripheral blood mononuclear cells. *Clinical and Experimental Immunology* 2005; 140:478-490.
15. Bogaert D, Thompson C, Trzcinski K, Malley R and Lipsitch M. The role of complement in innate and adaptive immunity to pneumococcal colonization and sepsis in a murine model. *Vaccine* 2010; 28(3): 681-685.
16. Tarr AW, Urbanowicz RA and Ball JK. The Role of Humoral Innate Immunity in Hepatitis C Virus Infection. *Viruses* 2012; 4:1-27.